

# Model-based characterisation of antibiotic activity in the presence of immune response: towards improved preclinical-to-clinical translation

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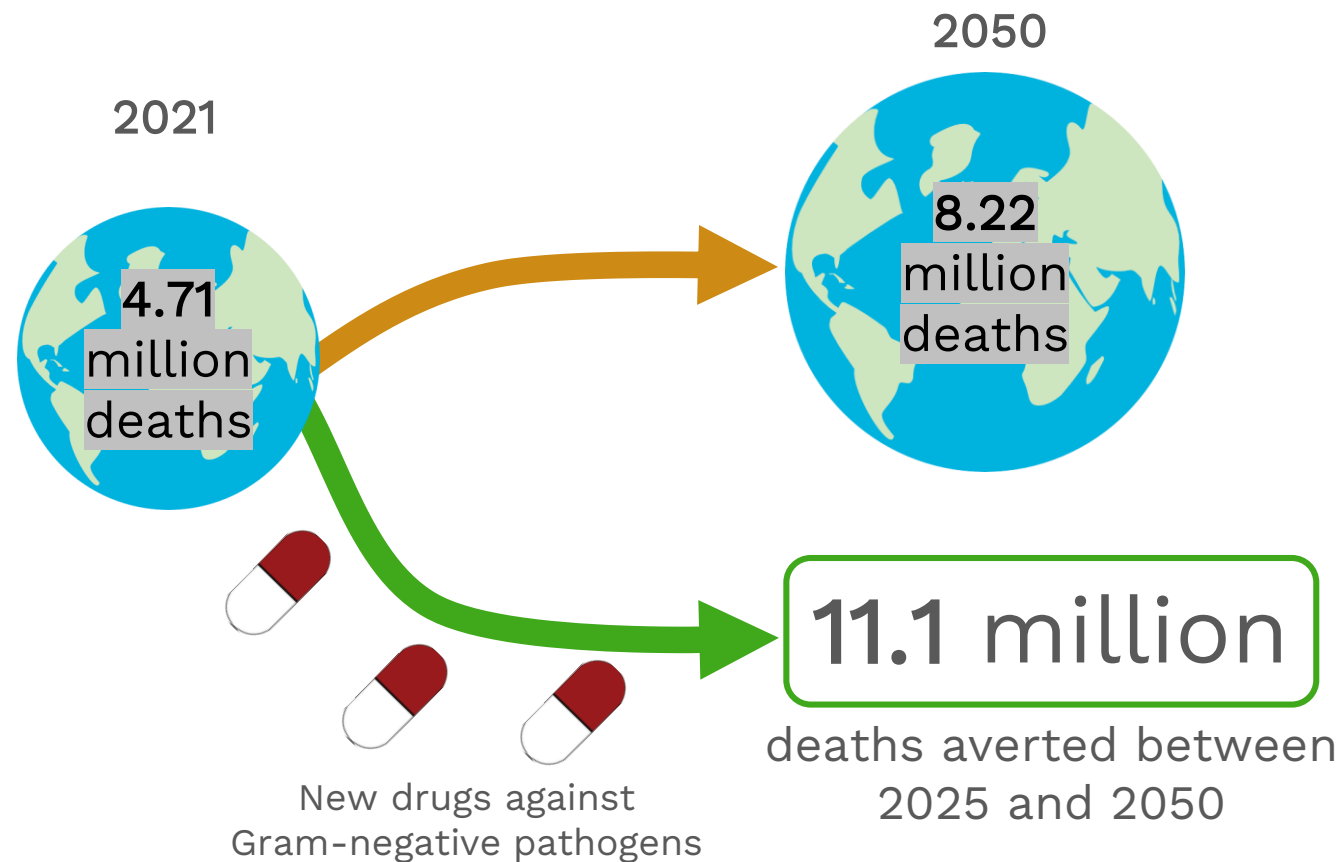
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# Antimicrobial resistance (AMR): a global threat

Global deaths per year associated with AMR<sup>1</sup>:



→ New antibiotics are needed!

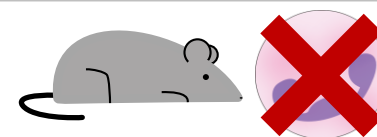


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# Preclinical evaluation of antibiotic PKPD



*In vitro* time-kill



*In vivo*

Neutropenic mouse thigh/lung infection



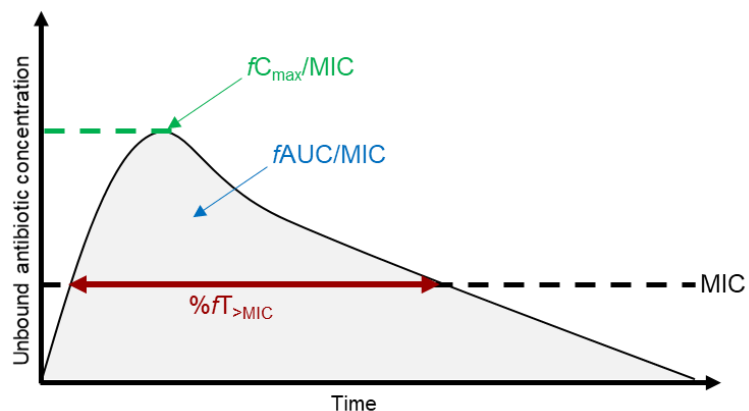
**Clinical use?**

- Bacteria not quantifiable in patients  
→ Reliance on preclinical data



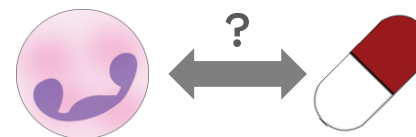
**Translation?**

- PK/PD indices: standard approach
- Limitations: single time point, summary metrics

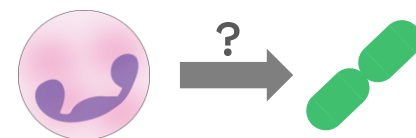


**Immune response?**

Preclinical settings are neutropenic:



interactions?



quantitative killing?

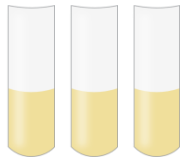


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# Evaluated antibiotics

## Afabicin

- Fatty acid synthesis inhibitor (FabI enzyme)
- Phase II development (NCT02426918, NCT03723551)<sup>2</sup>
- Available data (drug development):



*In vitro* time-kill



Neutropenic mouse thigh infection



Immunocompetent mouse thigh infection

## Meropenem

- Carbapenem (inhibition of cell wall synthesis)
- Study design:



Neutropenic mouse lung infection



Intermediate suppression mouse lung infection

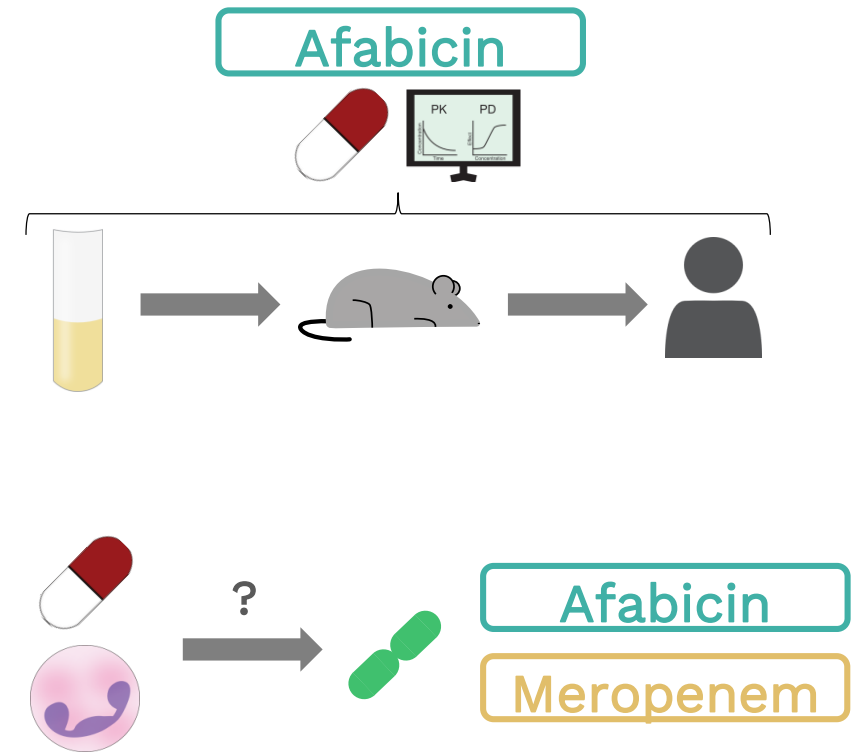


Immunocompetent mouse lung infection



# Aims

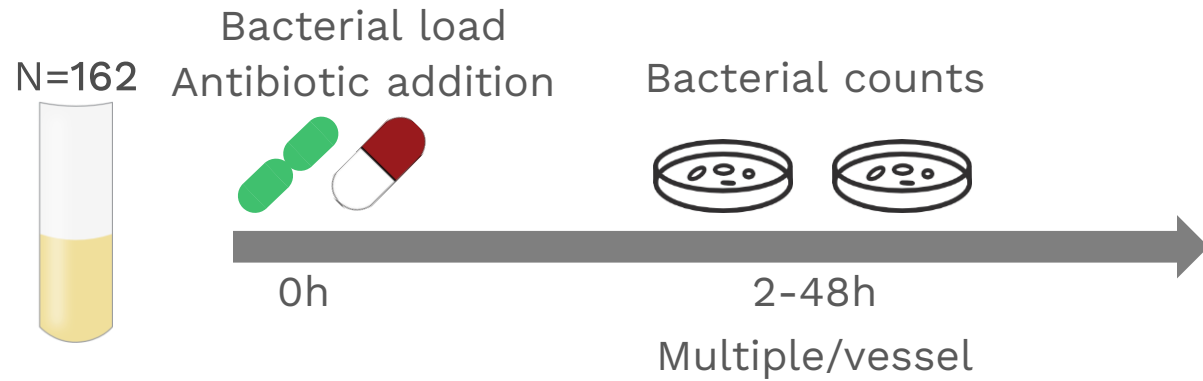
- To translate antibiotic activity from *in vitro* to *in vivo* and subsequently to clinical settings using PKPD modelling approaches
- To quantify the relative contribution of antibiotics and immune response to bacterial killing



# *In vitro* time-kill experiments



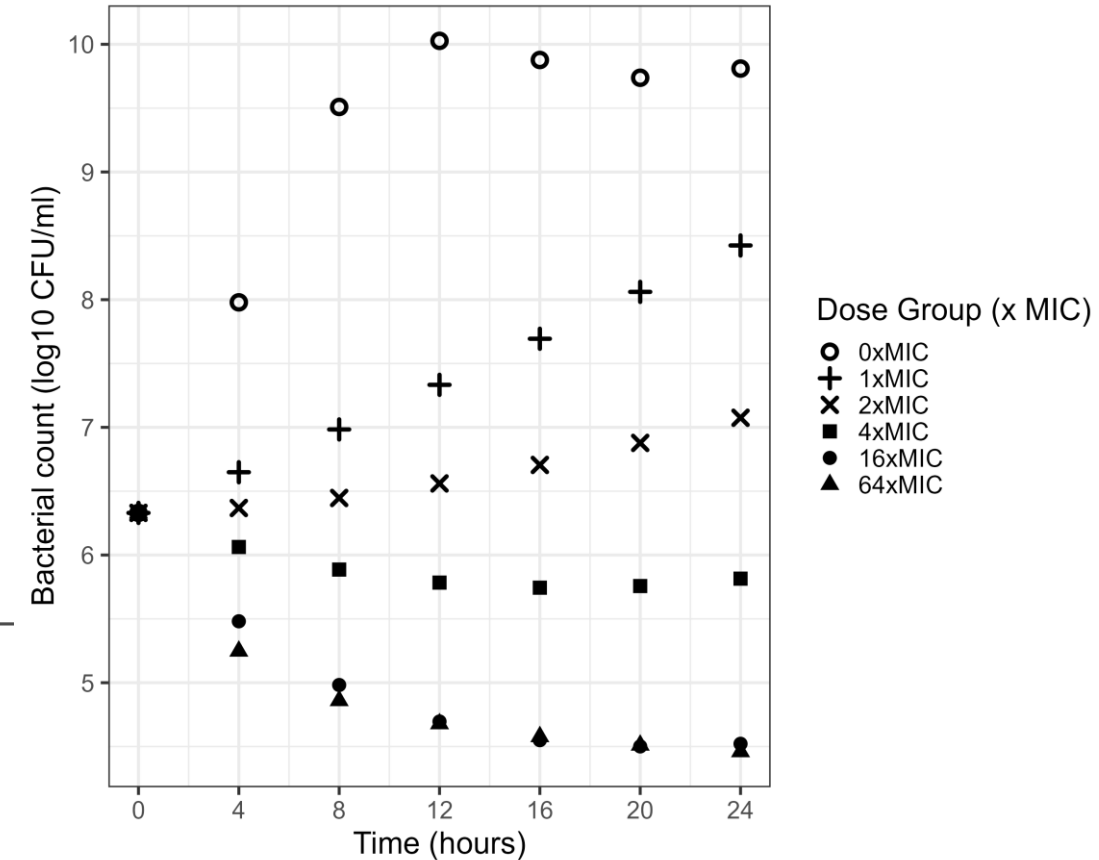
Bacterial dynamics  
over time



21 *Staphylococcus aureus* strains  
Minimum Inhibitory Concentration (MIC): 0.004 – 0.03 mg/L



Afabicin desphosphono (active metabolite)  
Static concentrations: 0.004 – 1 mg/L

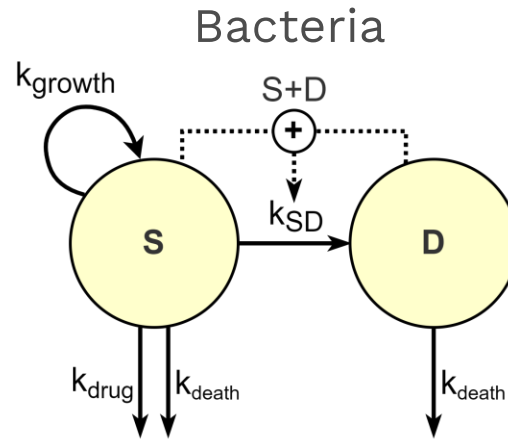


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# *In vitro* PKPD model



Susceptible bacteria (S):  
Growing, drug-susceptible

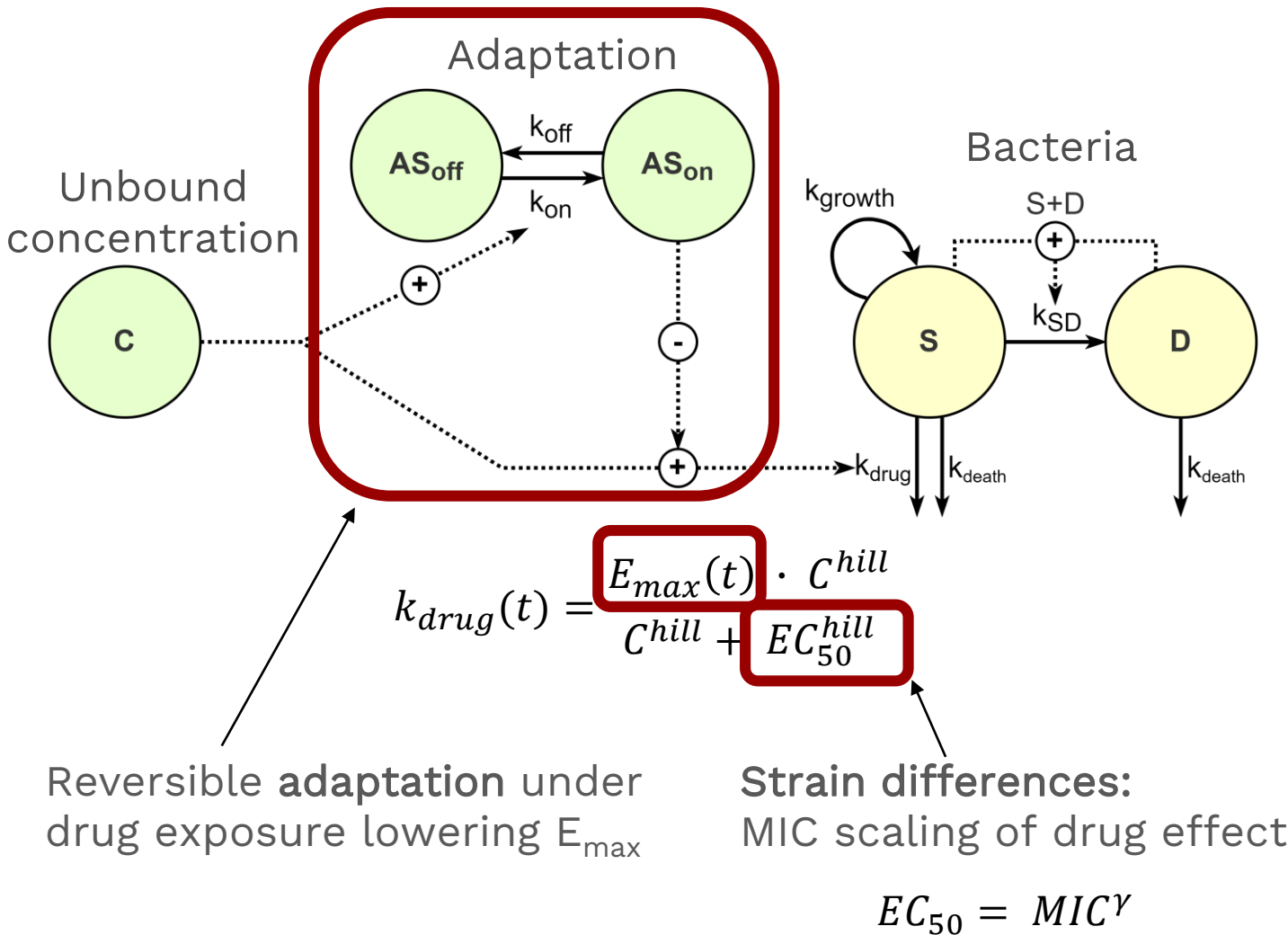


Dormant bacteria (D):  
Non-growing, non-drug-susceptible

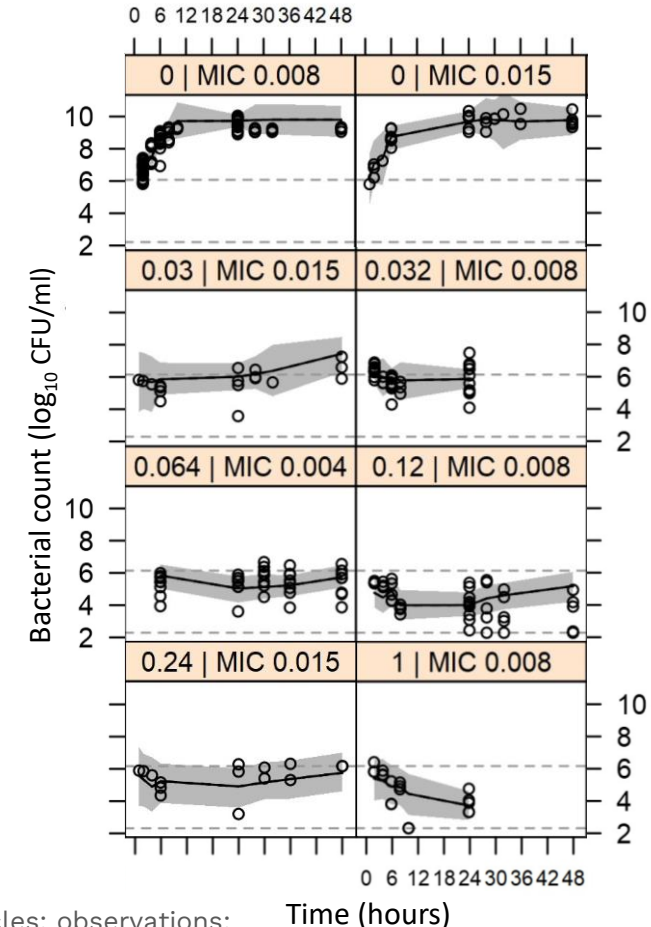
$S \rightarrow D$  transfer dependent on bacterial  
count and maximal system capacity



# In vitro PKPD model



Example subset of strains/concentrations:

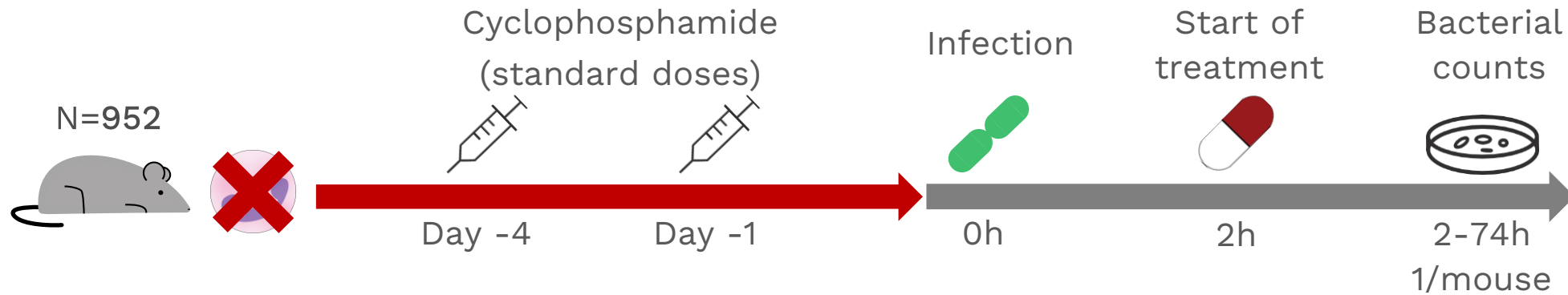



Circles: observations;  
solid lines (areas): median (95% CI) of model predictions;  
dashed lines: median start inoculum, limit of quantification




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# Neutropenic mouse thigh infection



 9 *Staphylococcus aureus* strains  
MIC: 0.004 – 0.06 mg/L  
8/9 strains not evaluated *in vitro* → translation across bacterial strains

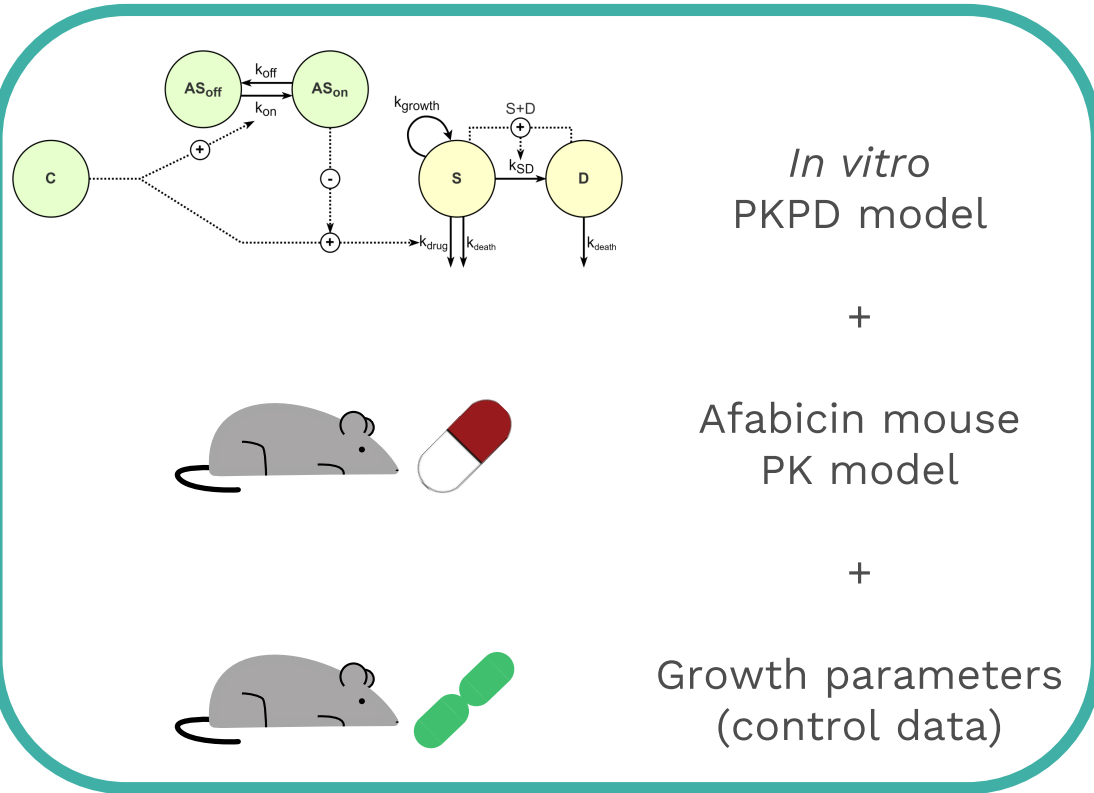
 Afabicin 0.01 – 190 mg/kg q6h



# Translation from *in vitro* to *in vivo*

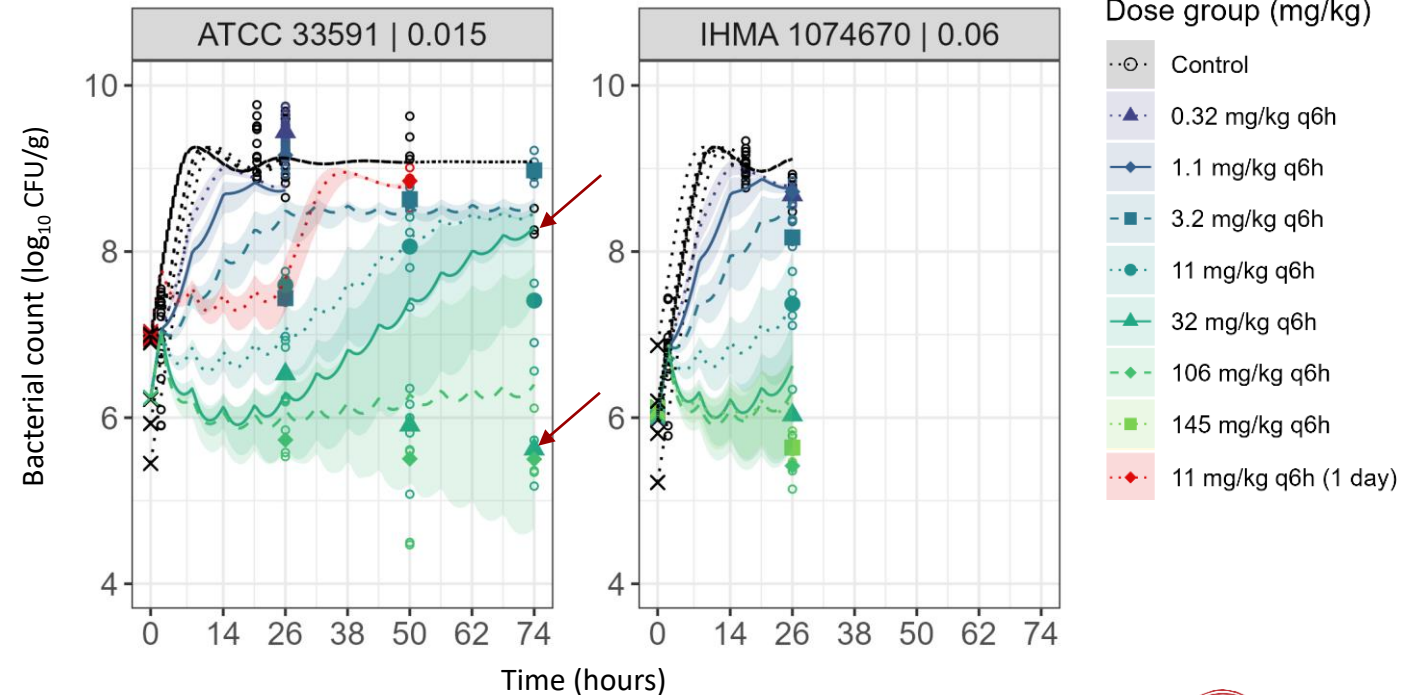


Can *in vivo* study outcomes be predicted using insights from *in vitro* studies?



## Predictions with PKPD parameter uncertainty

Example strains:



Symbols: median of observations; open circles: observations;  
lines: median of model predictions;  
areas: 95% confidence interval of predictions (uncertainty)



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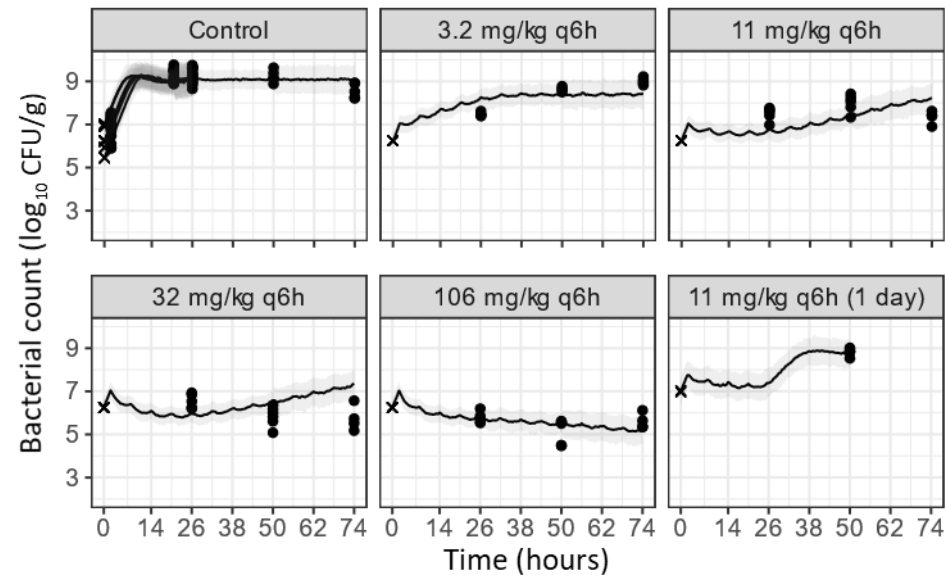
# Re-estimation with *in vivo* data



- Joint model estimation with *in vitro* and neutropenic mouse thigh infection data
- $EC_{u,50}$  38 to 45% lower *in vivo*, other drug effect parameters shared with *in vitro*

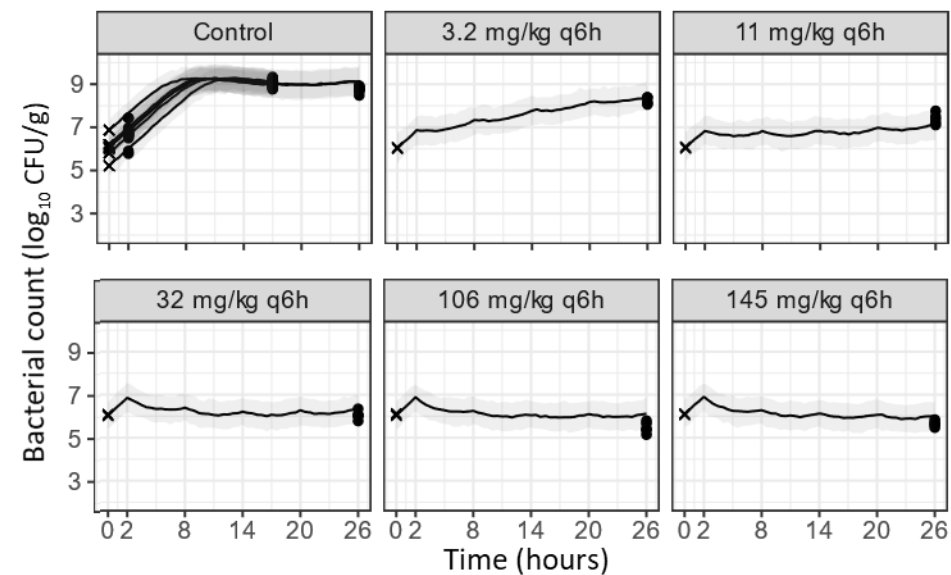
Example strains:

*S. aureus* ATCC33591 (MIC=0.015 mg/L)



Circles: observations; crosses: start inoculum;  
solid lines (areas): median (95% CI) of model predictions

*S. aureus* IHMA1074670 (MIC=0.06 mg/L)



All results in open-access article  
Saporta R, et al. Journal of Antimicrobial  
Chemotherapy. 2024;79(12):3150-59



# What about the immune system?

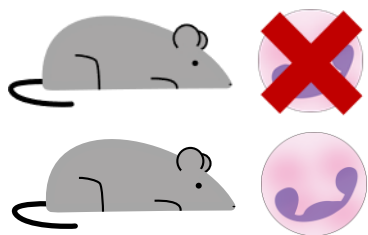
Afabicin

Meropenem



Influence of the immune system on antibiotic PK?

N=60



**Meropenem** PK study  
in infected mice  
(40 & 300 mg/kg)



Influence of the immune system on antibiotic effect?

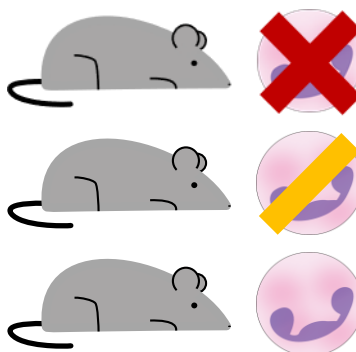
N=819



Inclusion of immunocompetent  
**afabicin** data in translation framework  
4 *S. aureus* strains  
(studied in neutropenic mice)



N=180



**Meropenem** efficacy study in  
mice with various immune states



40 or 300 mg/kg q4h



*K. pneumoniae* DSM116099  
MIC = 0.032 mg/L

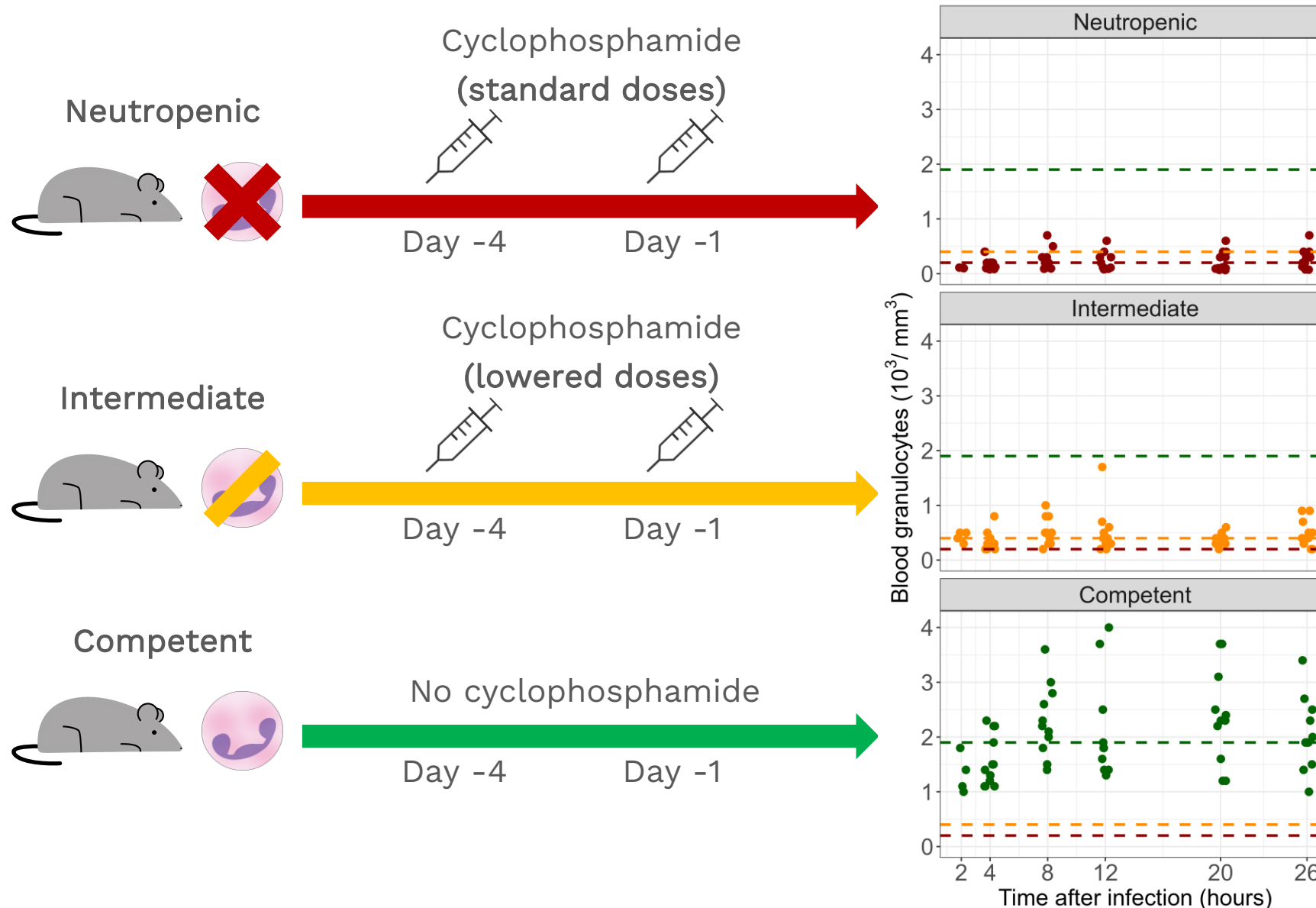


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# Immunocompetent mouse infection

Afabicin

Meropenem



Blood granulocytes in  
**meropenem** PD study  
(1 observation/mouse)

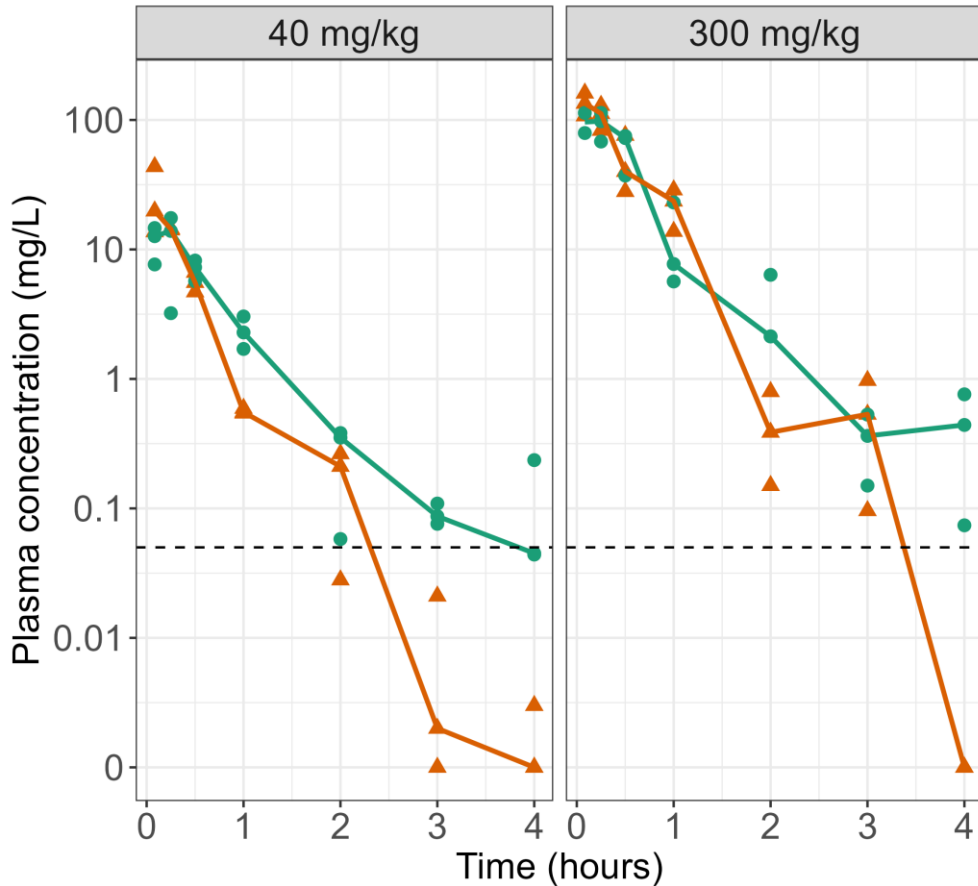
Circles: observations; dashed lines: medians of  
observations per immune status (all times)



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# Immune status influences meropenem PK

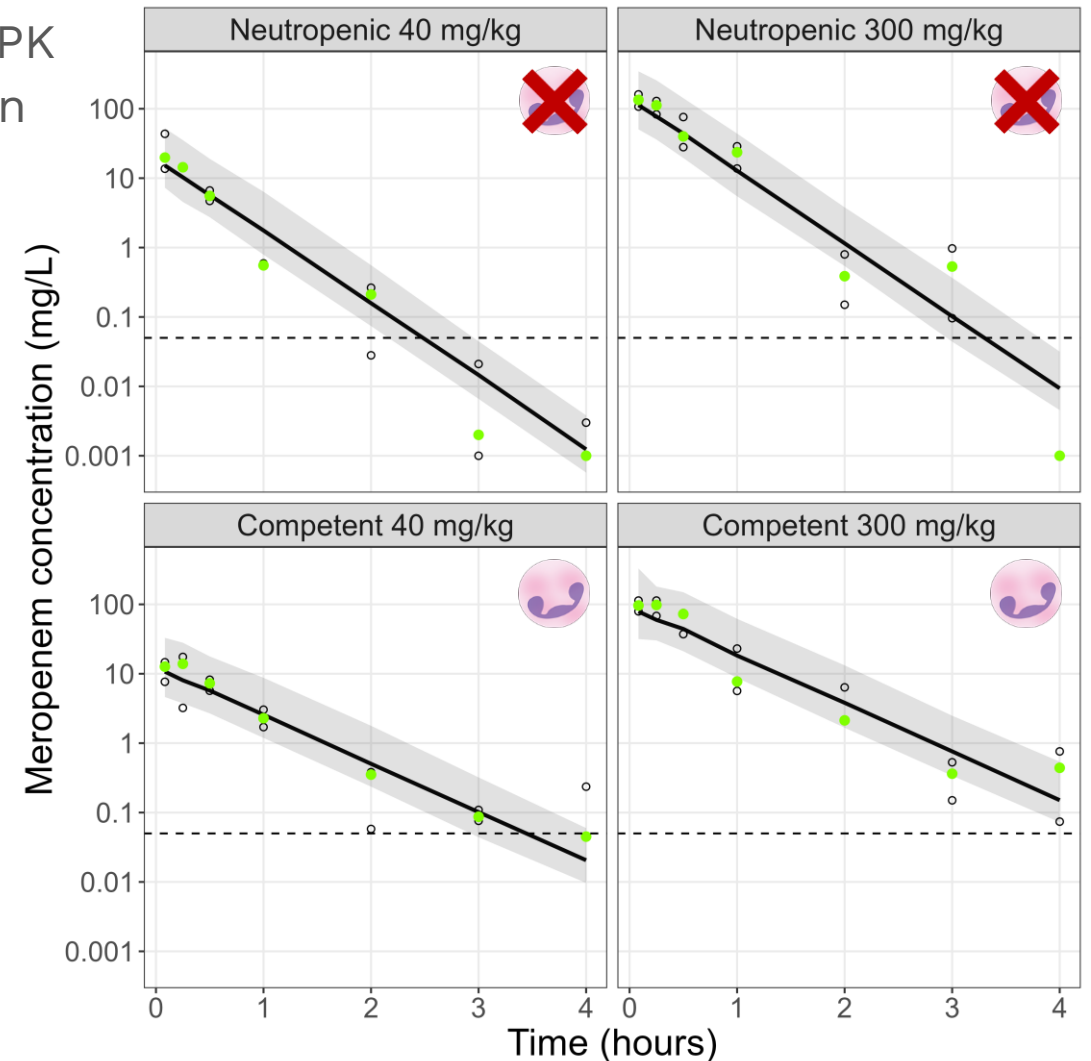
Meropenem



Immune status — Competent — Neutropenic

Symbols: observations; lines: median of observations;  
horizontal dashed line: limit of quantification (LOQ)

1 compartment PK  
≠ in distribution  
volume



(Green) circles: (median) observations;  
solid lines (areas): median (95% CI) of model predictions;  
horizontal dashed line: LOQ

# Immune response modelling

Afabicin

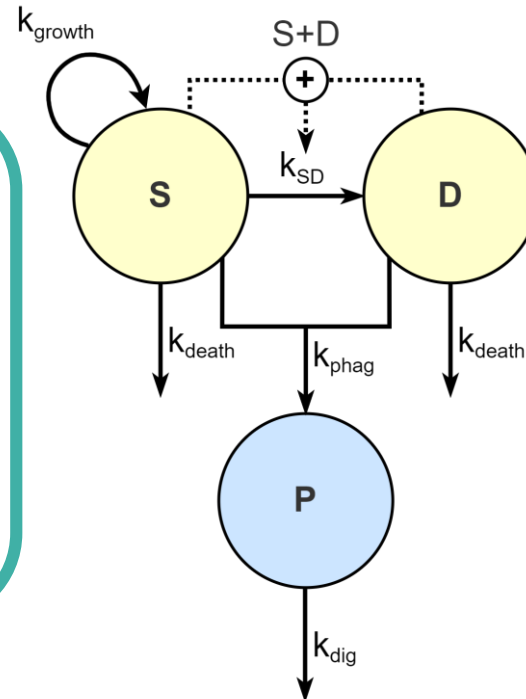
Meropenem

## Afabicin

- Neutrophil dynamics fixed from literature<sup>3</sup>
- Saturable phagocytosis rate:

$$k_{phag} = k_{phag,max} \cdot \left( 1 - \frac{P/N^H}{P/N^H + P/N_{50,phag}^H} \right)$$

Phagocytosis and digestion process



## Meropenem

- Immune status: covariate for  $k_{phag}$  and  $k_{dig}$



Neutropenic Intermediate Competent

$k_{phag} (h^{-1})$	0 (Fixed)*	0.187	0.321
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\*Non-differentiable from  $k_{growth}$

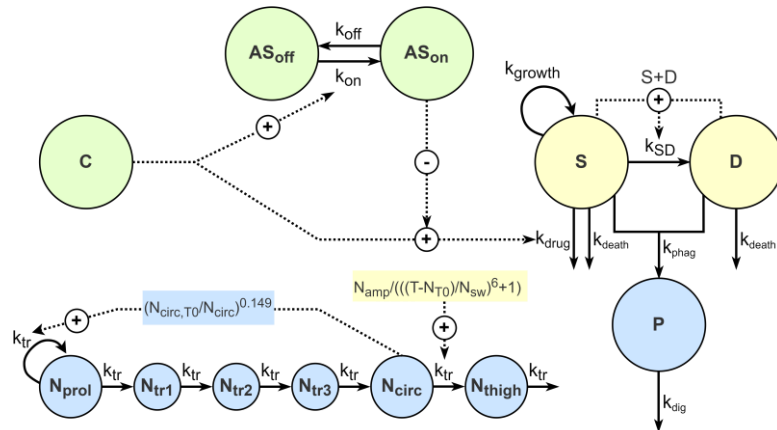
# Drug effect in presence of immune response

Afabacin

Meropenem

→ Lower contribution of antibiotics to bacterial killing in immunocompetent conditions

## Afabacin



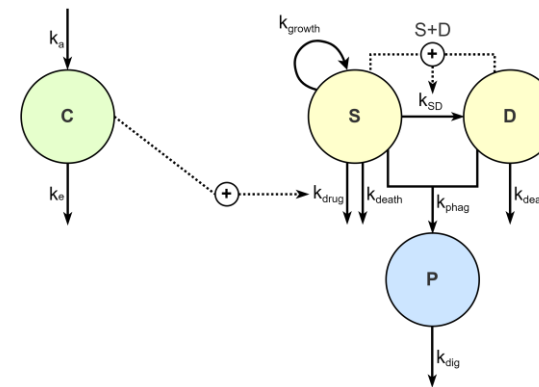
- Model for *in vitro* and neutropenic data expanded with killing by immune cells

- Reduced  $E_{max}$  and  $EC_{50}$  in immunocompetent vs neutropenic mice

$E_{max} (h^{-1})$	3.37	0.238
$EC_{u,50} (mg/L)$	0.264	0.074

Example for MIC=0.008 mg/L

## Meropenem



- Same  $E_{max}$  model parameters for all immune states

$E_{max} (h^{-1})$	0.922
$EC_{50} (mg/L)$	1.58

- Reduced effect attributed to a lower fraction of S bacteria due to phagocytosis

Available data did not support an intracellular drug effect on P bacteria



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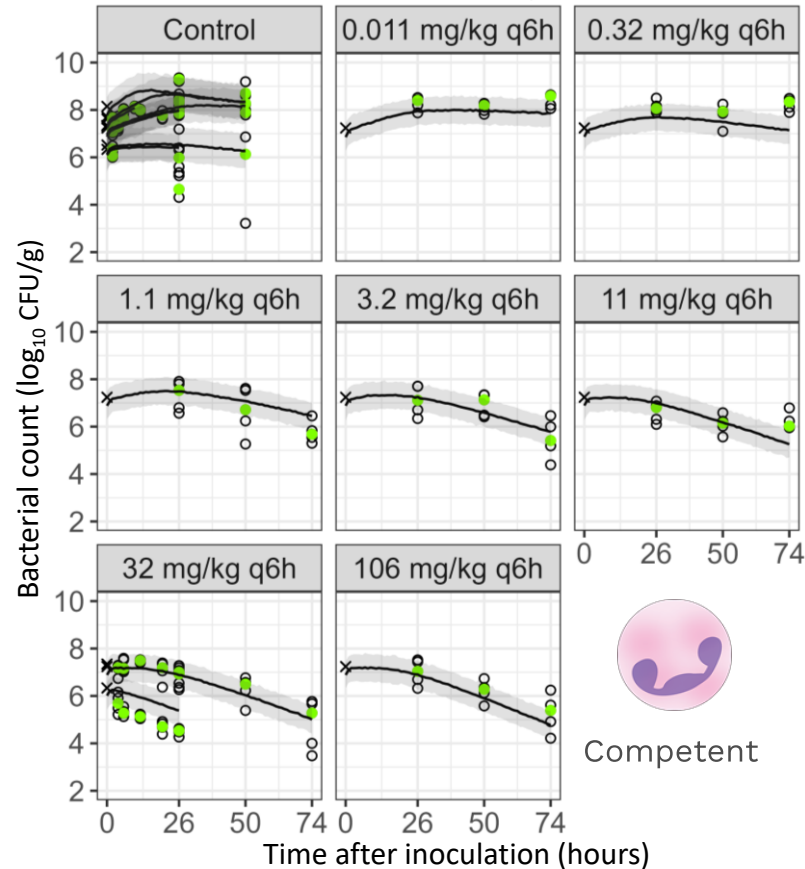
# The PKPD models described the time course of antibiotic effects and immune response

Afabicin

Meropenem

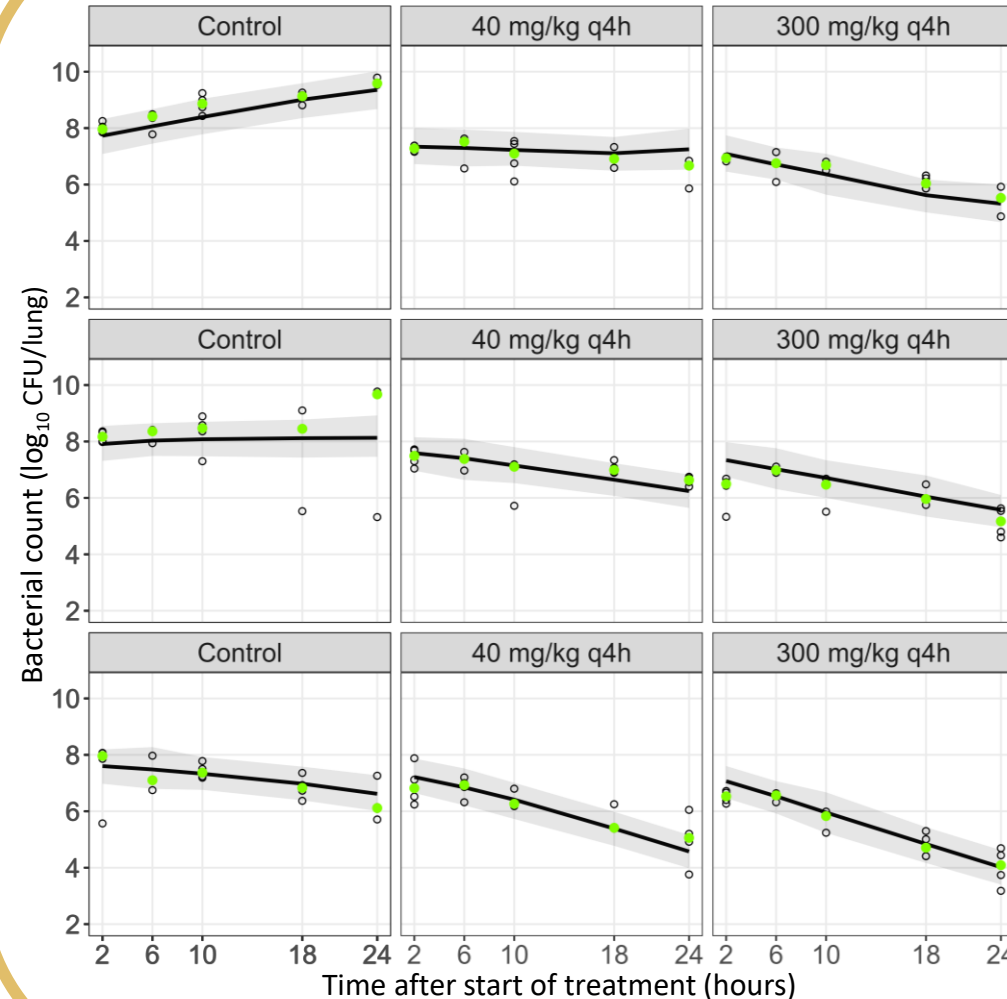
## Afabicin

Example strain: *S. aureus* ATCC33591  
MIC=0.015 mg/L



(Green) circles: (median) observations; crosses: inoculum;  
solid lines (areas): median (95% CI) of model predictions

## Meropenem



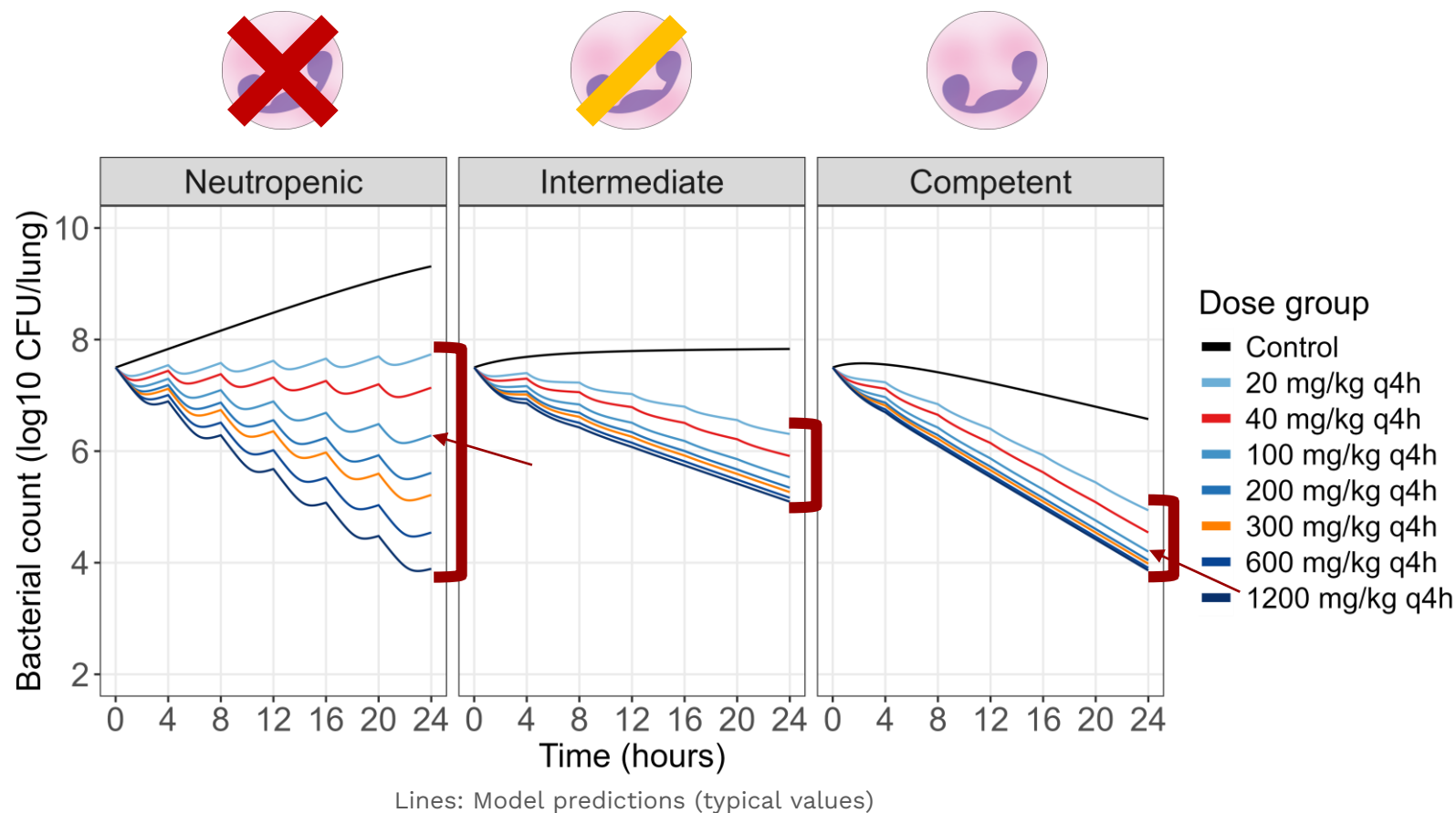
(Green) circles: (median) observations; solid lines (areas): median (95% CI)  
of model predictions; dashed lines: median of observations

MIC: Minimum Inhibitory Concentration

# Implications for dose-response relationships?

Meropenem

- Simulated meropenem dose-ranging study in mice with various immune states

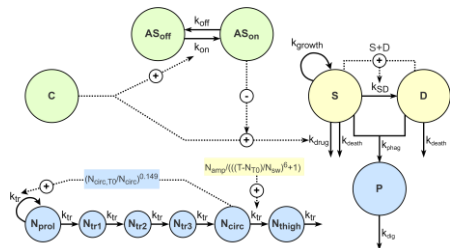


# Preclinical-to-clinical translation?



## Simulation settings

- Immunocompetent PKPD model
- Human PK model (Phase I data)
- Phase II dosing regimens**
- Covariates: sampled from distribution in NHANES database (adults)<sup>4</sup>
- Neutrophil levels:
  - $1.69 \times 10^6/\text{ml}$  (equivalent to mice)
  - $5 \times 10^6/\text{ml}$  (within normal range)

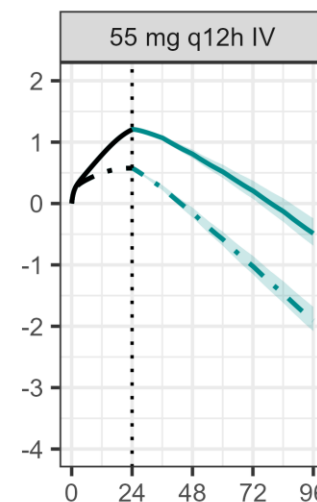
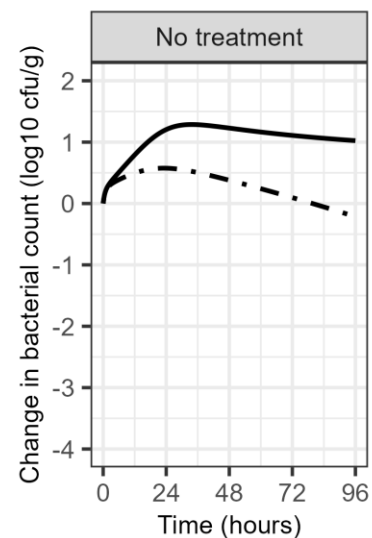


N=500  
per group

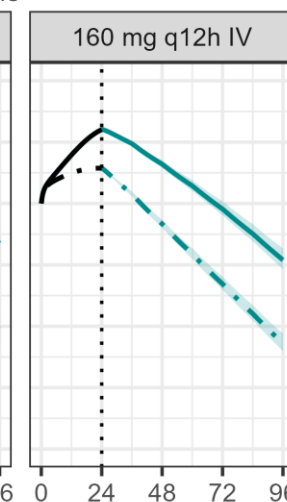
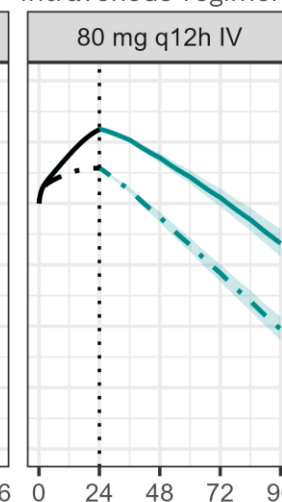
## Phase II results in ABSSSI<sup>2</sup>

- >90% early clinical responders
- Agreement of model-predicted bacterial dynamics and clinical response rates

ABSSSI: acute bacterial skin and skin structure infections



### Intravenous regimens

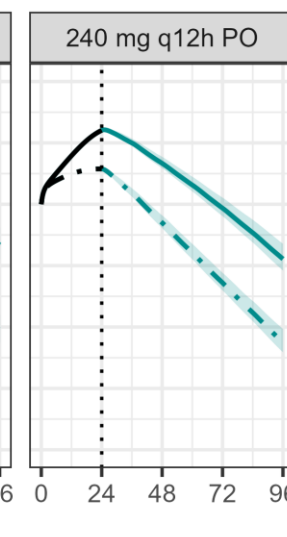
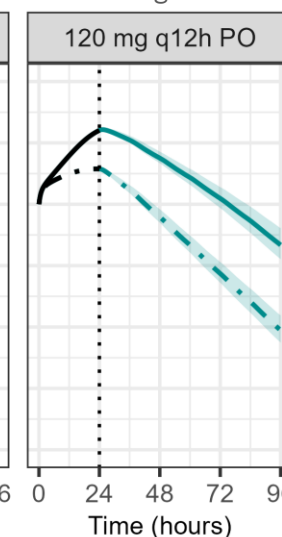
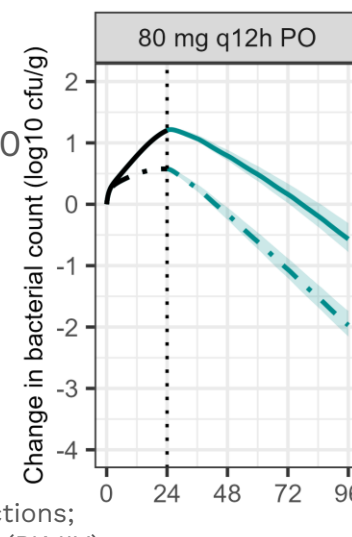


Example strain:  
*S. aureus* IHMA1074670  
MIC = 0.06 mg/L

Baseline neutrophils ( $10^6/\text{ml}$ )



### Oral regimens



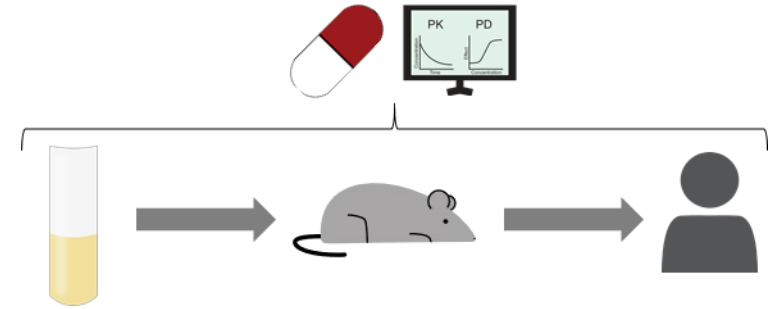
Lines: median of model predictions;  
areas: 90% prediction interval (PK IIV);  
vertical dashed line: start of treatment

<sup>2</sup>Wittke F, et al. Antimicrobial agents and chemotherapy 2020;64,10: e00250-20

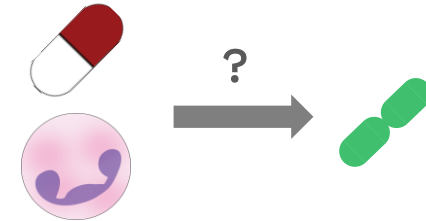
<sup>4</sup>Centers for Disease Control and Prevention. National Center for Health Statistics. National Health and Nutrition Examination Survey Data 08/2021-08/2023.

# Conclusions

Model-based approaches demonstrated an ability to translate antibiotic PKPD across bacterial strains and experimental settings



Antibiotic PK and/or efficacy were impacted by the immune system, leading to potential differences in dose-response



→ Quantifying the time course of bacterial killing by the antibiotic and the immune system with model-based approaches may improve translation and dose selection



# Acknowledgements



UPPSALA  
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Pharmacokinetics & Pharmacodynamics Research Group  
Pharmacometrics Research Group



Division of Infectiology



Translational Medicine Department



ITN members

## Funding:



Marie Skłodowska-Curie Actions



Swedish  
Research  
Council



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